

sufficiently different spectra). In the NMR experiment, one "average" environment for the ketone was found due to exchange of the ketone between the two media, one with water and one without. In the infrared the two environments are separable.

The results presented here give a picture of micelles which is compatible with current views.^{3,4} Water penetrates almost everywhere into micelles; there is a region (assumed to be the central core) completely devoid of water. Probe molecules are able to move in and out of the different environments. Further work is in progress using this methodology to probe the structure of other systems.

Acknowledgment. Thanks are due to Dr. U. Köhler for his advice on the synthesis of 7-oxo-octanoic acid and to Dr. J. Villalain for a preprint of his work.⁸

The Influence of a Carboxylate Group on the Rate of O-Acylation of 2-Hydroxymethylimidazoles by a Strained Amide

K. I. Skorey, V. Somayaji, and R. S. Brown*

Department of Chemistry, University of Alberta
Edmonton, Alberta, Canada T6G 2G2

Received March 1, 1988

During the course of all serine protease¹ catalyzed ester and amide hydrolyses, the Ser-OH group of the catalytic triad (Asp CO₂⁻-HisIm-SerOH) becomes transiently acylated. Although this suggests common hydrolytic pathways, continuing studies with serine proteases (SPases) indicate subtle substrate-dependent diversities. These include differing sites of initial acylation² and different numbers of protons in flight in the rate-limiting step.³ The role of the essential Asp CO₂⁻ component has not been generally resolved. Two possibilities, general base enhancement of the imidazole basicity and electrostatic stabilization of imidazolium, have been favored.¹ Evidence exists that enzymes inhibited with species approximating the initial tetrahedral intermediate maintain an Asp CO₂⁻-H-Im⁺-His H-bond, thus supporting the electrostatic role.⁴ Even so, it is possible that different substrates may recruit different levels of involvement of the various catalytic components. Nevertheless, the wide-spread occurrence of this catalytic triad suggests a considerable mechanistic advantage to the enzymes that employ it.

If such an arrangement leads to obvious acceleration, it is surprising how few studies have directly addressed the ability of a triad to facilitate O-acylation in a small molecule. A number of reports deal with the reaction of amino alcohols with esters⁵ or reactive amides.^{5b,6} A smaller number deal with the interaction

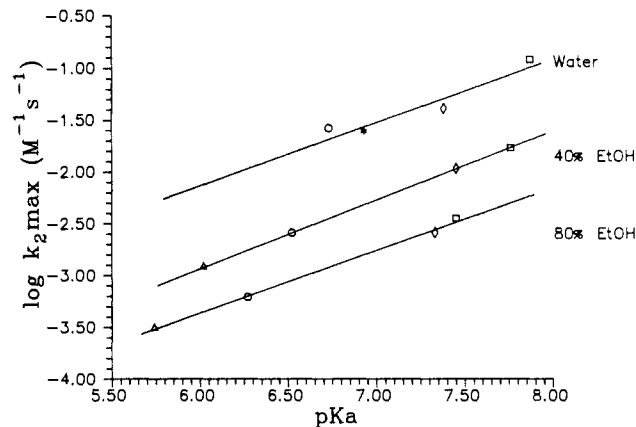


Figure 1. Brønsted plots of the maximal second-order rate constant (k_2^{\max}) versus pK_a^{Im} for **1a** (\diamond), **1b** (Δ), **1c** (\circ), **1d** ($*$), and **1e** (\square) with twisted amide **2** in H₂O ($\beta = 0.6 \pm 0.1$), 40% ($\beta = 0.66 \pm 0.03$) and 80% (0.60 ± 0.02) (v/v) EtOH/H₂O, $T = 25^\circ\text{C}$, $\mu = 0.2$ (KCl).

Table I. pK_a^{Im} Values for **1a-c,e** Determined by Potentiometric Titration at 25°C , $\mu = 0.2$ (KCl) in H₂O, 40% and 80% (v/v) EtOH/H₂O^a

compd	pK_a^{Im}		
	H ₂ O	40% EtOH/H ₂ O ^b	80% EtOH/H ₂ O ^b
1a	7.38 \pm 0.05	7.45 \pm 0.02	7.33 \pm 0.02
1b	6.18 \pm 0.02	6.02 \pm 0.02	5.74 \pm 0.02
1c	6.73 ^c	6.52 \pm 0.02	6.27 \pm 0.02
1e	7.88 \pm 0.03	7.76 \pm 0.02	7.45 \pm 0.02

^aAverages of duplicate measurements. ^bAdjusted pK_a values according to pH = (meter reading) - 0.09 (40% EtOH/H₂O) or pH = (meter reading) - 0.2 (80% EtOH/H₂O): Bates, R. G.; Paabo, M.; Robinson, R. A. *J. Phys. Chem.* **1963**, *67*, 1833. ^cEiki, T.; Kawada, S.; Matsushima, K.; Mori, M.; Tagaki, W. *Chem. Lett.* **1980**, 997.

of CO₂⁻ and imidazole during acyl transfer to H₂O⁷ (which is more properly a model for the deacylation of SPases). Apparently the preliminary reports of Bender et al.⁸ are the only published ones employing a model of the triad in acylation, in this case by *m*- and *p*-*tert*-butylphenylacetate.

We have shown that the direct O-acylation of **1c** and **d** by **2**⁹ proceeds to **3** via the process shown in eq 1.⁶ Herein we report an incremental study of the effect of the remote carboxylate in **1a** on the analogous process.

Shown in Figure 1 are Brønsted plots ($\log k_2^{\max}$ versus imidazole pK_a^{Im})¹¹ for reaction of **1a-e** with **2** in H₂O, 40% and 80% v/v EtOH/H₂O. (pK_a^{Im} and k_2^{\max} values are given as Supplementary Material.) Several common features are of note. (1) The $\log k_2$ values¹¹ plateau above pK_a^{Im} indicating the basic form is active. (2) The reaction product in all cases is CH₂-O-acylated as judged by ¹H NMR, IR, and mass spectral data (see Supplementary Material). (3) With no CH₂OH group present, the k_2^{\max} values

(1) For pertinent reviews of the structural and mechanistic properties of this class of enzymes see: (a) Walsh, C. *Enzymatic Reaction Mechanisms*; W. H. Freeman: San Francisco, CA, 1979; pp 56-97. (b) Fersht, A. *Enzyme Structure and Mechanism*, 2nd ed.; W. H. Freeman: San Francisco, CA, 1985. (c) Dugas, H.; Penney, C. *Bioorganic Chemistry*; Springer Verlag: New York, 1981; pp 208-226. (d) Schowen, R. L. In *Principles of Enzyme Activity*; Liebman, J. F.; Greenberg, A. Eds.; VCH Publishers Inc. U.S.A., Vol. 9, in press.

(2) (a) Hubbard, C. D.; Shoupe, T. S. *J. Biol. Chem.* **1977**, *252*, 1633. (b) Hubbard, C. D.; Kirsch, J. F. *Biochemistry* **1972**, *11*, 2483. (c) Quinn, D. M.; Elrod, J. P.; Ardis, R.; Friesen, P.; Schowen, R. L. *J. Am. Chem. Soc.* **1980**, *102*, 5358.

(3) (a) Stein, R. L.; Strimpler, A. M. *J. Am. Chem. Soc.* **1987**, *109*, 4387, and references therein. (b) Venkatasubban, K. S.; Schowen, R. L. *CRC Crit. Rev. Biochem.* **1984**, *17*, 1.

(4) (a) Abeles, R. H.; Liang, T. C. *Biochemistry* **1987**, *26*, 7603. (b) Bone, R.; Shenvi, A. B.; Kettner, C. A.; Agard, D. A. *Ibid.* **1987**, *26*, 7609.

(5) (a) For leading references see footnote 3 of ref 6. (b) Cram, D. J.; Lam, P. Y.-S.; Ho, S. P. *J. Am. Chem. Soc.* **1986**, *108*, 839. (c) Cram, D. J.; Katz, H. E.; Dicker, I. B. *J. Am. Chem. Soc.* **1984**, *106*, 4987.

(6) Somayaji, V.; Skorey, K. I.; Brown, R. S.; Ball, R. G. *J. Org. Chem.* **1986**, *51*, 4866.

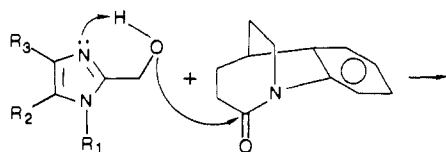
(7) (a) Komiyama, M.; Bender, M. L. *Bioorg. Chem.* **1977**, *6*, 13. (b) Komiyama, M.; Bender, M. L.; Utaka, M.; Takeda, A. *Proc. Natl. Acad. Sci. U.S.A.* **1977**, *74*, 2634. (c) Mallick, I. M.; D'Souza, V. T.; Yamaguchi, M.; Lee, J.; Chalabi, P.; Gadwood, R. C.; Bender, M. L. *J. Am. Chem. Soc.* **1984**, *106*, 7252. (d) Rogers, G. A.; Bruice, T. C. *J. Am. Chem. Soc.* **1974**, *96*, 2473. (e) Komiyama, M.; Breaux, E. J.; Bender, M. L. *Bioorg. Chem.* **1977**, *6*, 127.

(8) (a) D'Souza, V. T.; Hanabusa, K.; O'Leary, T.; Gadwood, R. C.; Bender, M. L. *Biochem. Biophys. Res. Commun.* **1985**, *129*, 727. (b) D'Souza, V. T.; Bender, M. L. *Acc. Chem. Res.* **1987**, *20*, 146. (c) The above two (ref 8a and b) report an interesting cyclodextrin with an attached pendant imidazolyl benzoate facilitating the hydrolysis of *m*- and *p*-*tert*-butylbenzoate. Evaluation of the extent of cooperativity is difficult at present since full experimental details and controls are not yet reported.

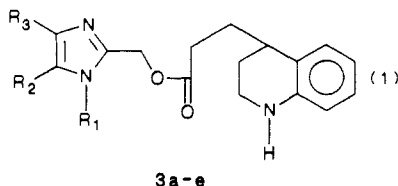
(9) Somayaji, V.; Brown, R. S. *J. Org. Chem.* **1986**, *51*, 2676.

(10) All new imidazoles had satisfactory elemental and spectroscopic (IR, ¹H NMR, MS) analyses. The product esters from **1a-e** and **2** were characterized by ¹H NMR, exact mass, and IR (Supplementary Material).

(11) Second-order rate constants (k_2) for **1a-e** with **2** were evaluated from slopes of the $\log k_{\text{obsd}}$ versus $[1a-e]$ plots at different pH values. The maximal second-order rate constant (k_2^{\max}) was calculated from $k_2 = k_2^{\max} K_a^{\text{Im}} / ([H^+] + K_a^{\text{Im}})$.



- 1a: $R_1 = R_2 = H$, $R_3 = C(CH_3)_2CO_2^-$
 1b: $R_1 = R_2 = H$, $R_3 = C(CH_3)_2CO_2Me$
 1c: $R_1 = R_2 = R_3 = H$
 1d: $R_1 = CH_3$, $R_2 = R_3 = H$
 1e: $R_1 = H$, $R_2 = R_3 = CH_3$



lie ~ 8 –100 fold (depending on the example⁶) below the Brønsted line indicating this group is required for maximal activity.⁶ For example, if the CH_2OH unit in **1a** is replaced by CH_2OCH_3 , the k_2^{max} value drops from $(4.06 \pm 0.05) \times 10^{-2}$ to $(5.90 \pm 0.45) \times 10^{-3} M^{-1} s^{-1}$.

Since **1a** lies on the same Brønsted lines as defined by **1b–e**, the CO_2^- unit in this system is incapable of providing another mechanism for O-acylation¹² other than that simply dependent upon imidazole basicity. A fundamentally different or enhanced pathway such as CO_2^- acting as a general base (on N–H) in concert with the imidazole–HO interaction is expected to produce an upward deviation from the line.

Since this is not observed, at first glance one might suggest there is no positive benefit of attachment of the CO_2^- in **1a**, but we believe the situation is more subtle. The pK_a^{im} values in Table I show the N basicity of **1a** is greater than that of comparison ester **1b**, and a corresponding increase in k_2^{max} is observed. In EtOH/H₂O media of reduced polarity,¹³ activity of all the imidazole alcohols is reduced as expected, and there is a noticeable drop in the pK_a^{im} of **1b–e**. However, the electrostatic and/or H-bonding stabilization in the zwitterionic form of **1a** counteracts the general medium-induced reduction in N basicity¹⁴ thereby enhancing the CH_2OH nucleophilicity in relation to that of **1b–e**.

In summary, the main benefit of the anionic pendant in **1a** is an electrostatic one which is manifested more prominently in media of reduced polarity. To term this system a “model” for the acylation of SPases invites comparison with the enzyme which may not be justified given the unorthodox structure of the acylating agent⁶ and perhaps nonoptimal orientation of the functional groups in **1a**.¹⁵ Rather, we prefer to view the system as a small molecule demonstration of an electrostatic role for CO_2^- –Im in direct CH_2OH acylation. Nevertheless we note that in addition to other possibilities,¹ a similar electrostatic role has been suggested for the Asp CO_2^- –His section of the triad in the SPases.^{4,16}

Acknowledgment. We thank the University of Alberta and Natural Sciences and Engineering Research Council of Canada for financial support.

(12) The “trimethyl” lock engendered by the isobutyryl moiety compresses the carboxylate into a close H-bonding relationship with the imidazole as noted by Rogers and Bruce.^{7d} In the latter study, a negligible effect was noted in the deacylation relative to the corresponding carboxylic ester.

(13) Åkerlöf, G. *J. Am. Chem. Soc.* **1932**, *54*, 4125.

(14) For a discussion of the effect of alcohol/H₂O mixtures on amine, acid, and amino acid pK_a 's see: (a) Merle, M.; Douhéret, G.; Dondon, M.-L. *Bull. Soc. Chim. Fr.* **1966**, *Ser. 5*, 159. (b) Grunwald, E.; Berkowitz, B. J. *J. Am. Chem. Soc.* **1951**, *83*, 4939. (c) Chattopadhyay, A. K.; Lahiri, S. C. *Indian J. Chem.* **1977**, *15A*, 930.

(15) Gandour (Gandour, R. D. *Bioorg. Chem.* **1981**, *10*, 169) has described the orientation requirements for optimal general base activity of CO_2^- .

(16) The electrostatic role of Asp CO_2^- in enhancing the basicity of the imidazole in SPases has been discussed by the following: (a) Roberts, J. D.; Kanamori, K. *Proc. Natl. Acad. Sci. U.S.A.* **1980**, *77*, 3095. (b) Bachovchin, W. W. *Biochemistry* **1986**, *25*, 7751. (c) Bachovchin, W. W.; Roberts, J. D. *J. Am. Chem. Soc.* **1978**, *100*, 8041.

Supplementary Material Available: Tables of thermodynamic pK_a^{im} and k_2^{max} values for **1a–e** reacting with **2** and product study data (¹H NMR, IR, mass spectral) (3 pages). Ordering information is given on any current masthead page.

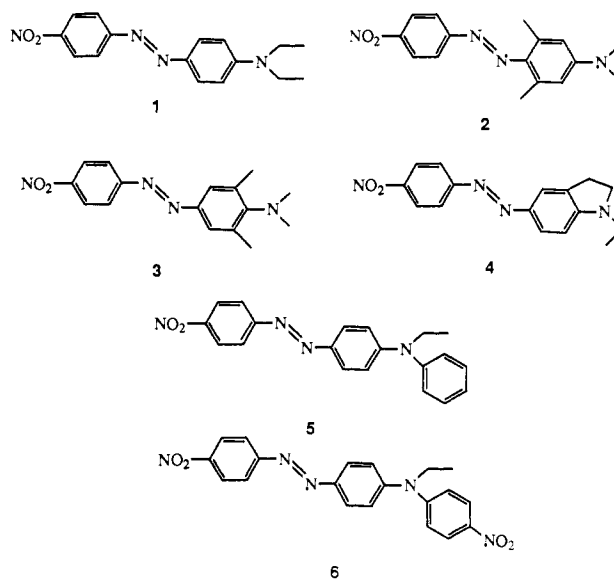
Solvent-Induced Mechanism Change in Charge-Transfer Molecules. Inversion versus Rotation Paths for the Z → E Isomerization of Donor–Acceptor Substituted Azobenzenes

Dong-Myung Shin and David G. Whitten*

Department of Chemistry, University of Rochester
 Rochester, New York 14627

Received March 4, 1988

The mechanism for the thermal Z → E isomerization of p-donor–p'-acceptor (“push–pull”) azobenzenes has generated considerable controversy.^{1–9} While reaction of azobenzene is generally accepted to proceed by an inversion mechanism, in which sp^2 – sp rehybridization of an azobenzene nitrogen affords isomerization via a semilinear transition state,^{10–15} we and others have noted coupling of donor and acceptor substituents in compounds such as 4-(diethylamino)-4'-nitroazobenzene (DENAB) (**1**) can



- (1) Asano, T.; Okada, T. *J. Org. Chem.* **1986**, *51*, 4454.
 (2) Asano, T.; Okada, T. *J. Org. Chem.* **1984**, *49*, 4387.
 (3) Marcandalli, B.; Liddo, L. P.-D.; Fede, C. D.; Bellobono, I. R. *J. Chem. Soc., Perkin Trans. 2* **1984**, 660.
 (4) Nishimura, N.; Kosako, S.; Sueishi, Y. *Bull. Chem. Soc. Jpn.* **1984**, *57*, 1617.
 (5) Nishimura, N.; Tanaka, T.; Asano, M.; Sueishi, Y. *J. Chem. Soc., Perkin Trans 2* **1986**, 1839.
 (6) Wildes, P. D.; Pacifici, J. G.; Irick, G., Jr.; Whitten, D. G. *J. Am. Chem. Soc.* **1971**, *93*, 2004.
 (7) Schanze, K. S.; Mattox, T. F.; Whitten, D. G. *J. Org. Chem.* **1983**, *48*, 2808.
 (8) Andersson, J. *J. Photochem.* **1983**, *22*, 245.
 (9) Sigman, M. E.; Leffler, J. E. *J. Org. Chem.* **1987**, *52*, 3123.
 (10) Talaty, E. R.; Fargo, J. C. *J. Chem. Soc., Chem. Commun.* **1967**, 65.
 (11) Harberfield, P.; Block, P. M.; Lux, M. S. *J. Am. Chem. Soc.* **1975**, *97*, 5840.
 (12) Asano, T.; Okada, T.; Shinkai, S.; Shigematsu, K.; Kusano, Y.; Manabe, O. *J. Am. Chem. Soc.* **1981**, *103*, 5161.
 (13) Rau, H.; Ludecke, E. *J. Am. Chem. Soc.* **1982**, *104*, 1616.
 (14) Asano, T.; Yano, T.; Okada, T. *J. Am. Chem. Soc.* **1982**, *104*, 4900.
 (15) Ljunggren, S.; Wettermark, G. *Acta Chem. Scand.* **1971**, *25*, 1599.